



SEMA

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**The non-canonical SOS-system of DNA repair and
mutagenesis in *Acinetobacter baumannii***

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Acinetobacter baumannii

- Gram-negative bacterium
- Nosocomial pathogen
- Resistant to most antibiotics



Atypical SOS response

The bacterial SOS system in *E. coli*

Global DNA damage response



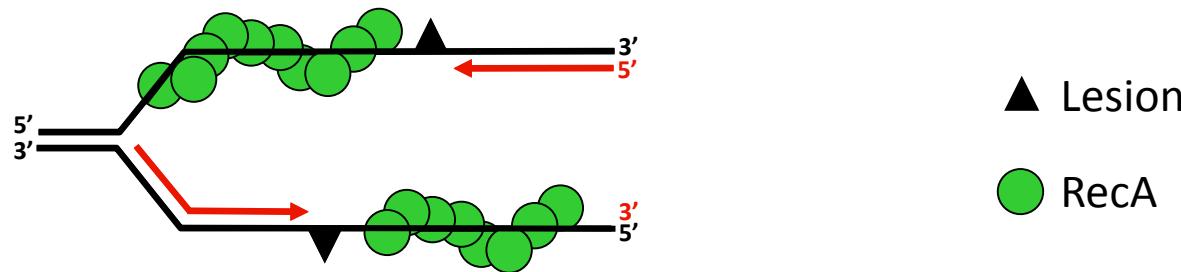
Repair of DNA lesions



Cell survival

The bacterial SOS system in *E. coli*

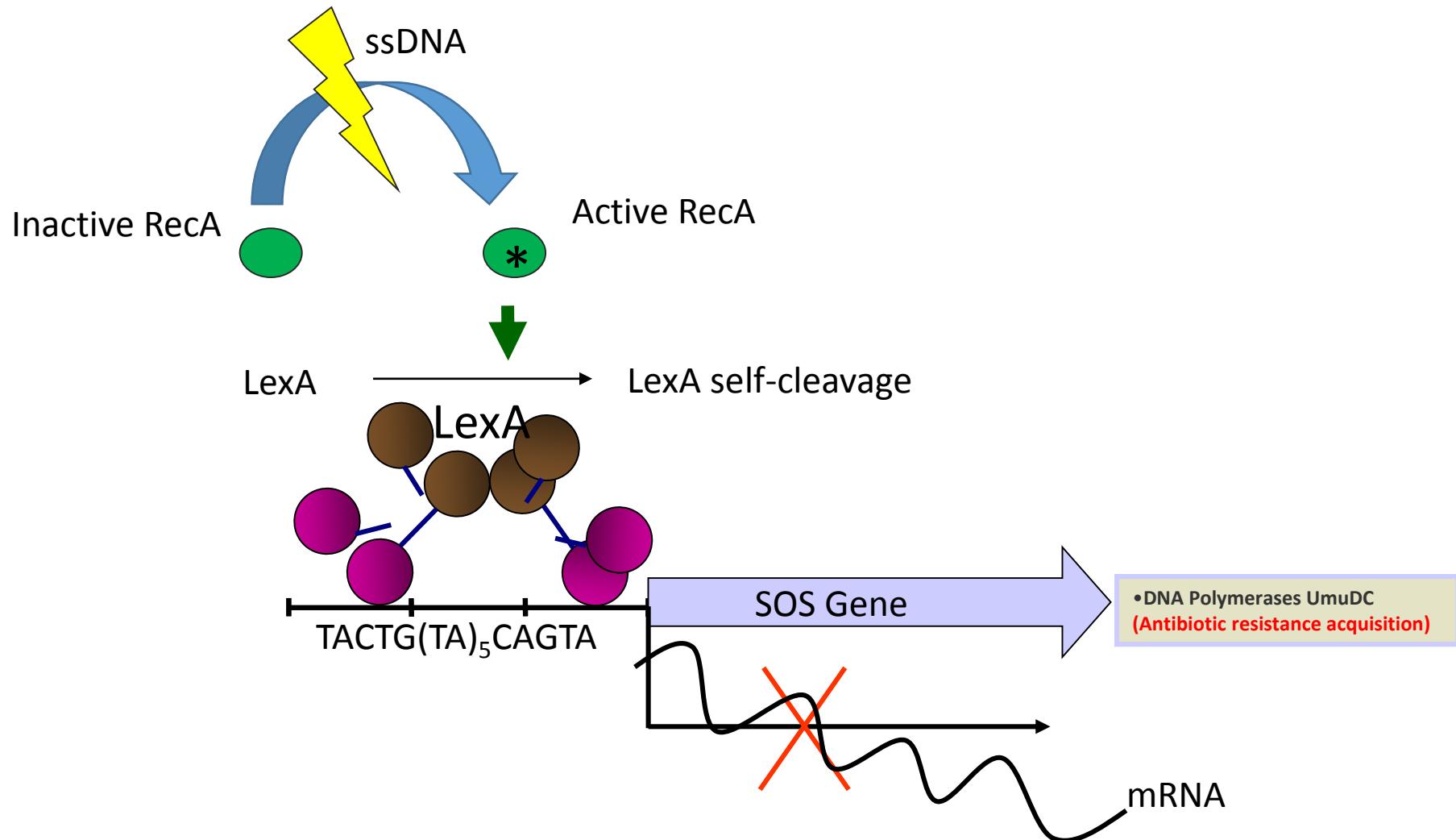
- DNA damage blocks DNA replication



- RecA forms a filament around these ssDNA regions becoming activated

Based on Little *et al.* Proc Natl Acad Sci U S A. 1980;77(6):3225-9.

The bacterial SOS system in *E. coli*



Based on Little et al. Proc Natl Acad Sci U S A. 1980;77(6):3225-9.

A*cinetobacter* spp.

- In *Acinetobacter* spp. no LexA homolog has been identified
- None of the promoters of genes of *Acinetobacter* spp. contain a known SOS sequence
- There is no damage-induced mutagenesis, with the remarkable exception of the opportunistic pathogens *Acinetobacter baumannii*, *Acinetobacter ursingii*, and *Acinetobacter beijerinckii*

Objectives

- I. Construction and characterization of a *recA* knock-out mutant of *A. baumannii*
- II. Analysis of the transcriptional response of *A. baumannii* to DNA-damaging agents
- III. Analysis of the DNA damage-induced mutagenesis in *A. baumannii*

Objective I

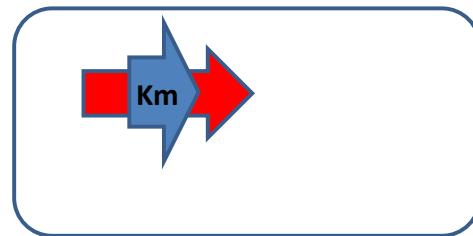
Characterization of a *recA* knockout
mutant of *A. baumannii*

The role of RecA in *A. baumannii*

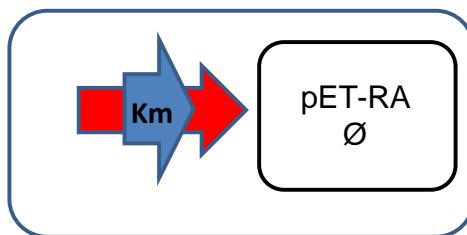
A. baumannii strains used in this work:



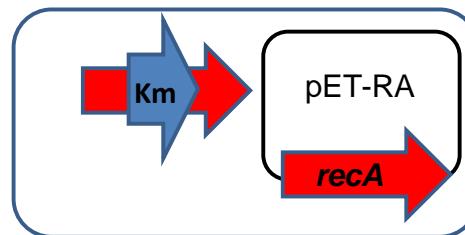
ATCC17978 (WT)



recA mutant

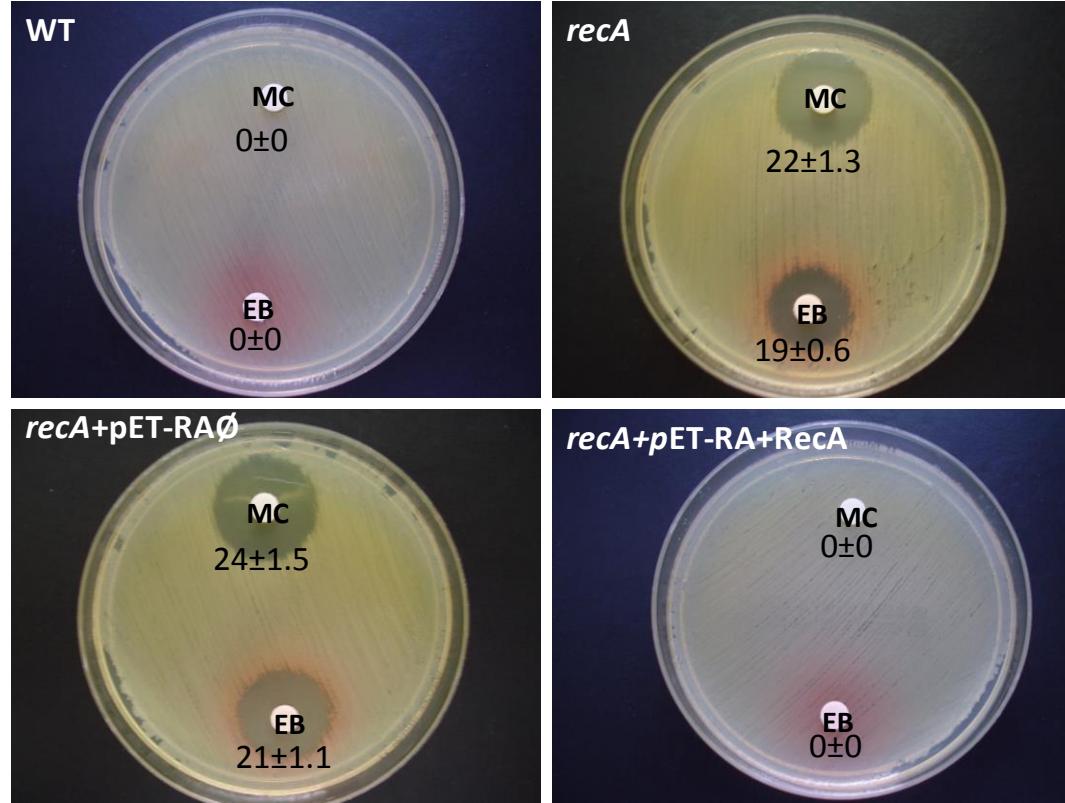


recA mutant + empty vector



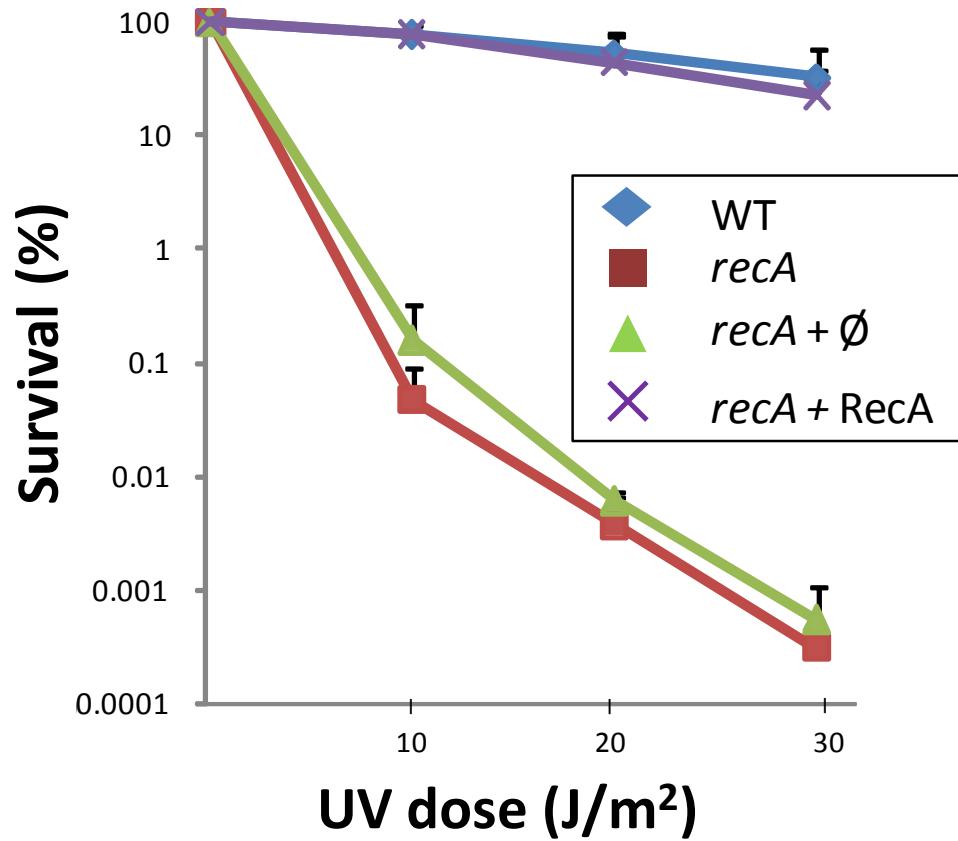
Complemented recA mutant

DNA damaging agents



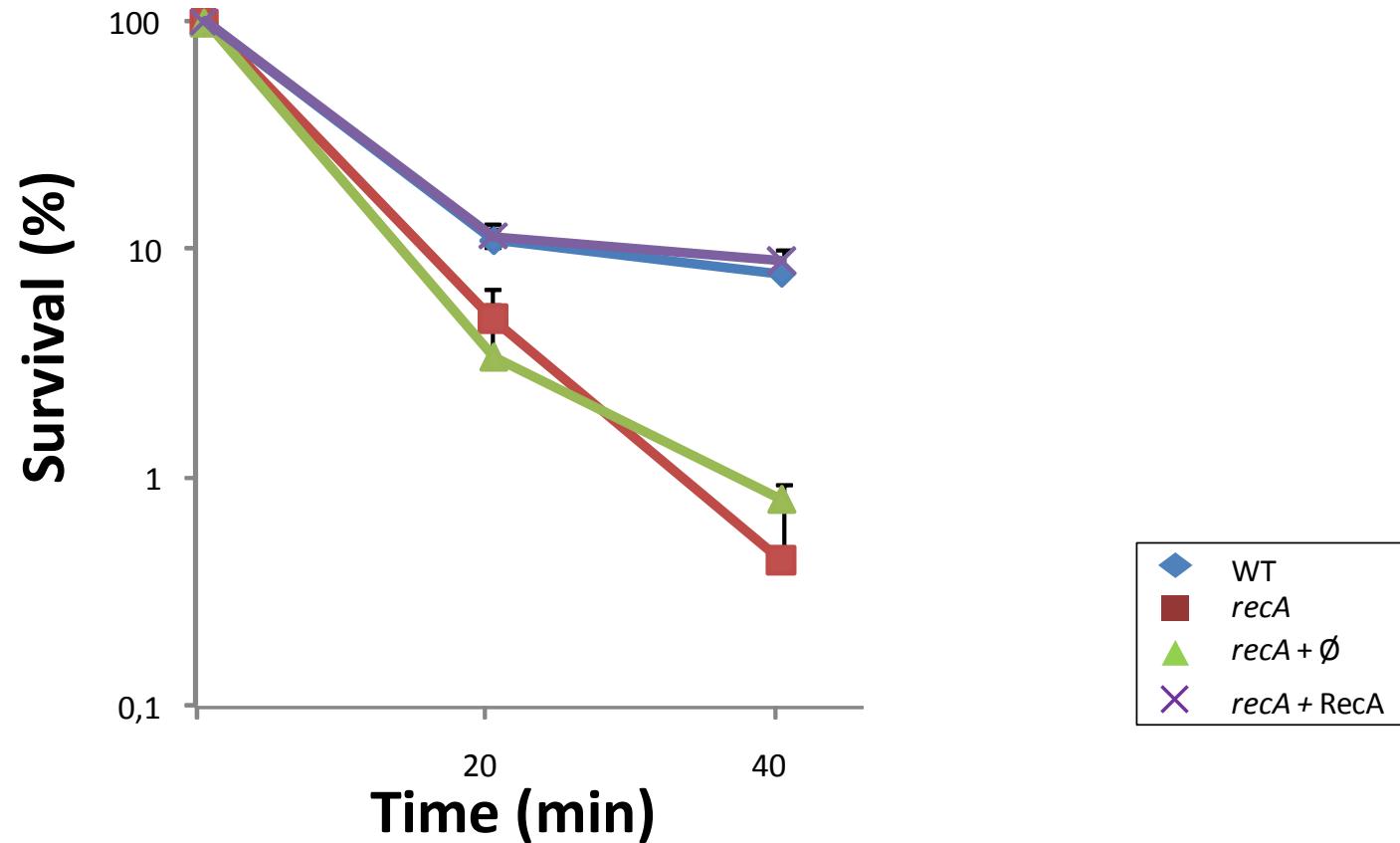
MC:MitomycinC; **EB:** Ethidium bromide
(Inhibition halos are indicated in mm)

DNA damaging agents: UV

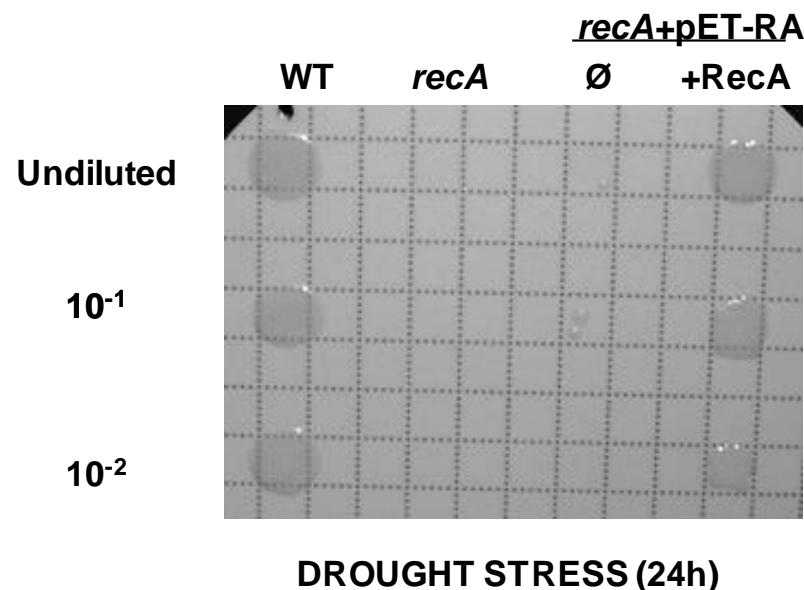
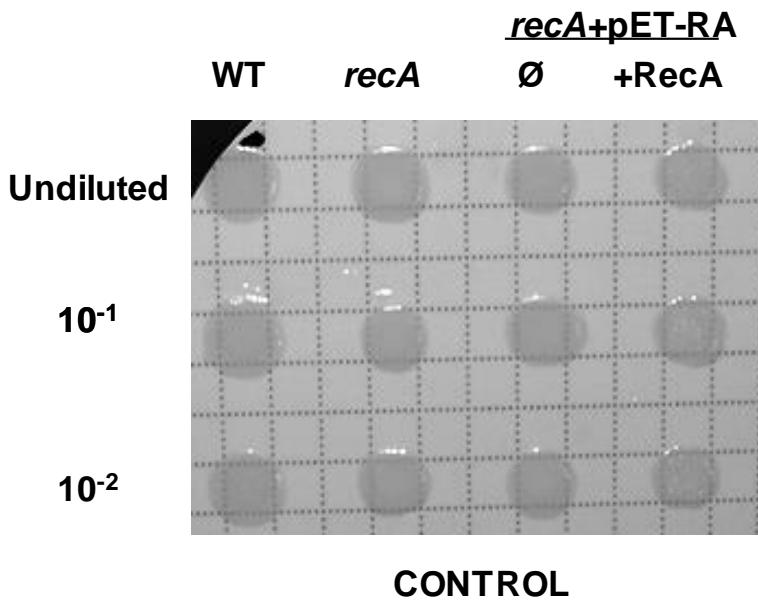


T

Thermal stress (55°C)

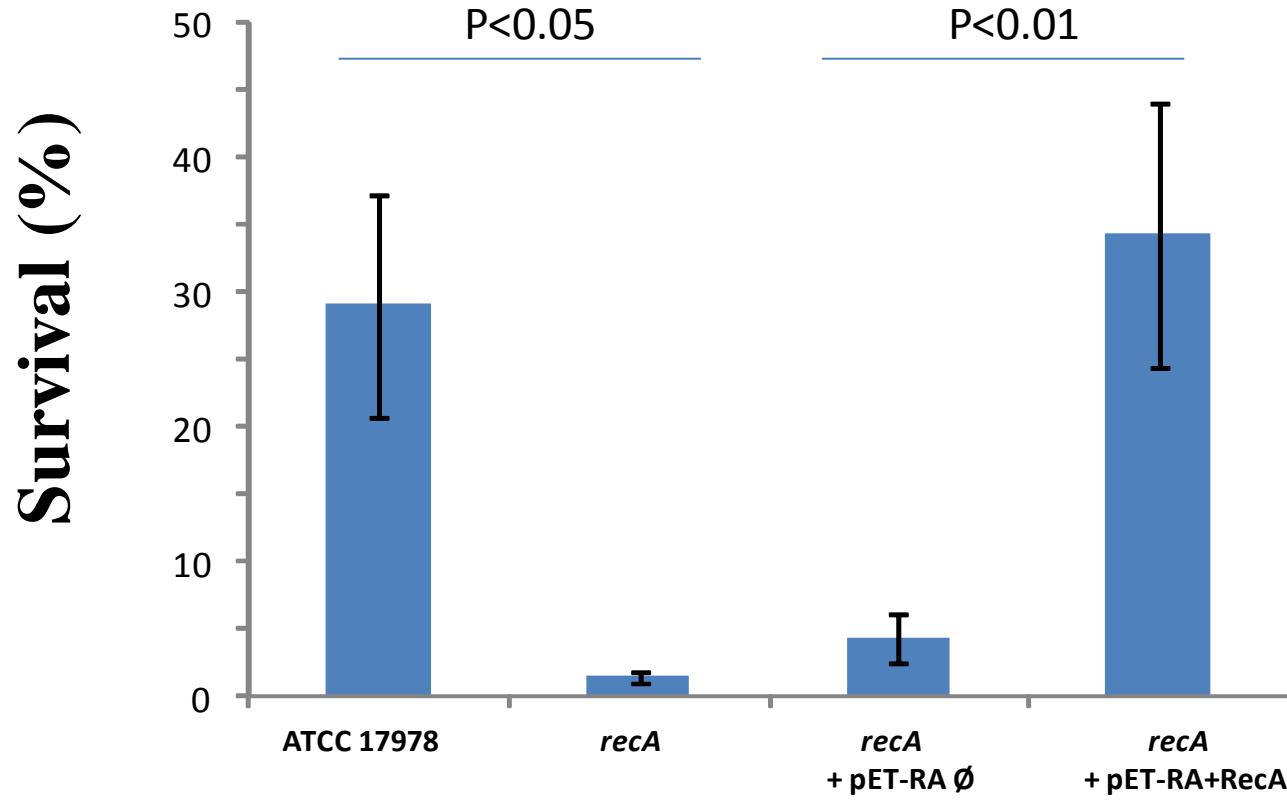


Desiccation



Desiccation resistance assays. Dilution series of exponentially growing cells (OD_{600} of 0.5) were spotted on sterile cellulose filters (0.45- μ m pore size; Millipore). The filters were either not dried (control) or dried for 24 h inside a sterile petri plate at 37°C before they were placed on tryptic soy agar (TSA) plates and incubated for 24 h at 37°C.

Virulence *in vitro* (macrophages)



Survival of the indicated strains of *A. baumannii* after 1-h incubation with macrophages (Cell line Raw 264.7).

Virulence *in vivo* (mice)

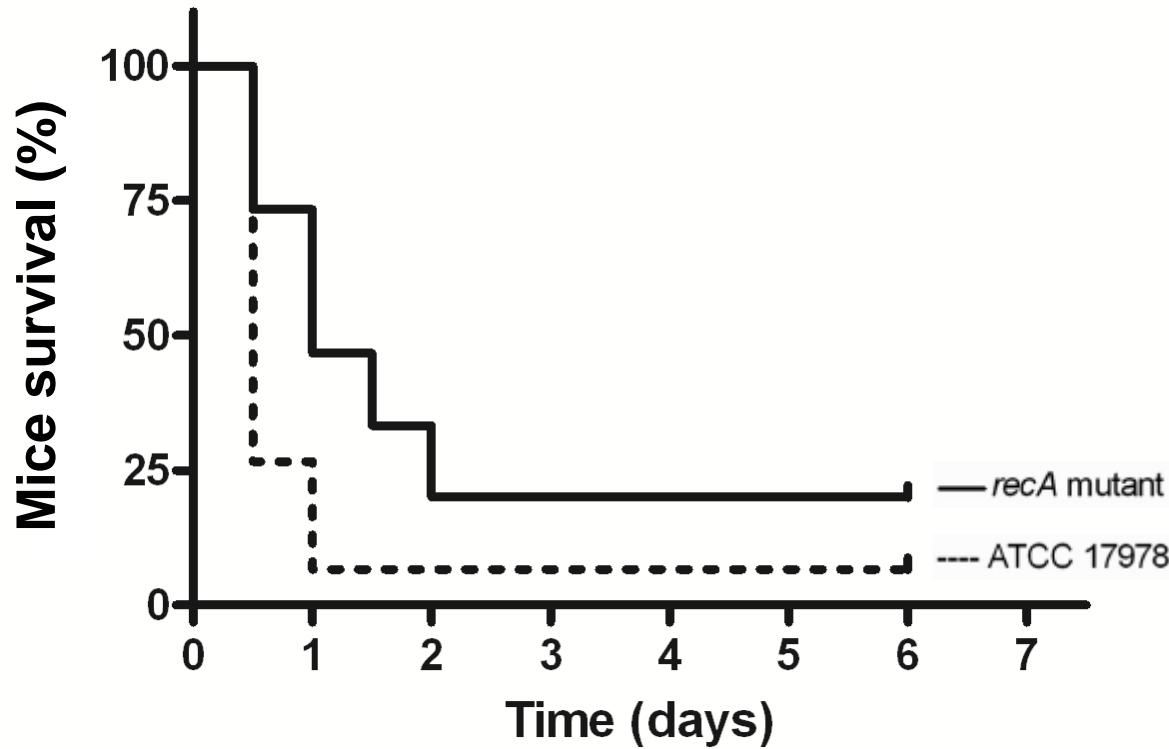


FIG. 7. Survival of mice ($n = 15$ per group) inoculated with the *A. baumannii* WT or *recA* mutant strain. Significant differences in survival were noted (log rank test, $P < 0.05$).

2×10^8 CFU/mouse

ICR (CD1) female mice (Harlan Interfauna Ibérica, Barcelona, Spain)

Antimicrobial susceptibilities

MICs of the antimicrobials used in this study for the indicated *A. baumannii* strains

Antibiotic	MIC of the indicated ATCC 17978 strain(µg/ml) ^a			
	Parent strain	<i>recA</i> mutant	<i>recA</i> pET-RAØ strain ^b	<i>recA</i> pET-RA-RecA strain
Piperacillin	32	12	12	24
Piperacillin/tazobactam	16	6	6	12
Amoxicillin/clavulanic acid	64	24	24	48
Imipenem	0.38	0.19	0.19	0.38
Ceftazidime	6	3	3	6
Cefotaxime	96	48	48	128
Colistin	0.5	0.25	0.25	0.38
Trimethoprim/sulfamethoxazole	4	0.75	0.75	6
Ciprofloxacin	0.25	0.016	0.016	0.25
Moxifloxacin	0.094	0.003	0.003	0.094
Erythromycin	4	4	4	6
Chloramphenicol	>256	>256	>256	>256
Tobramycin	0.38	0.38	0.38	0.38
Amikacin	1.5	1.5	1.5	2
Rifampin	4	4	NA	NA

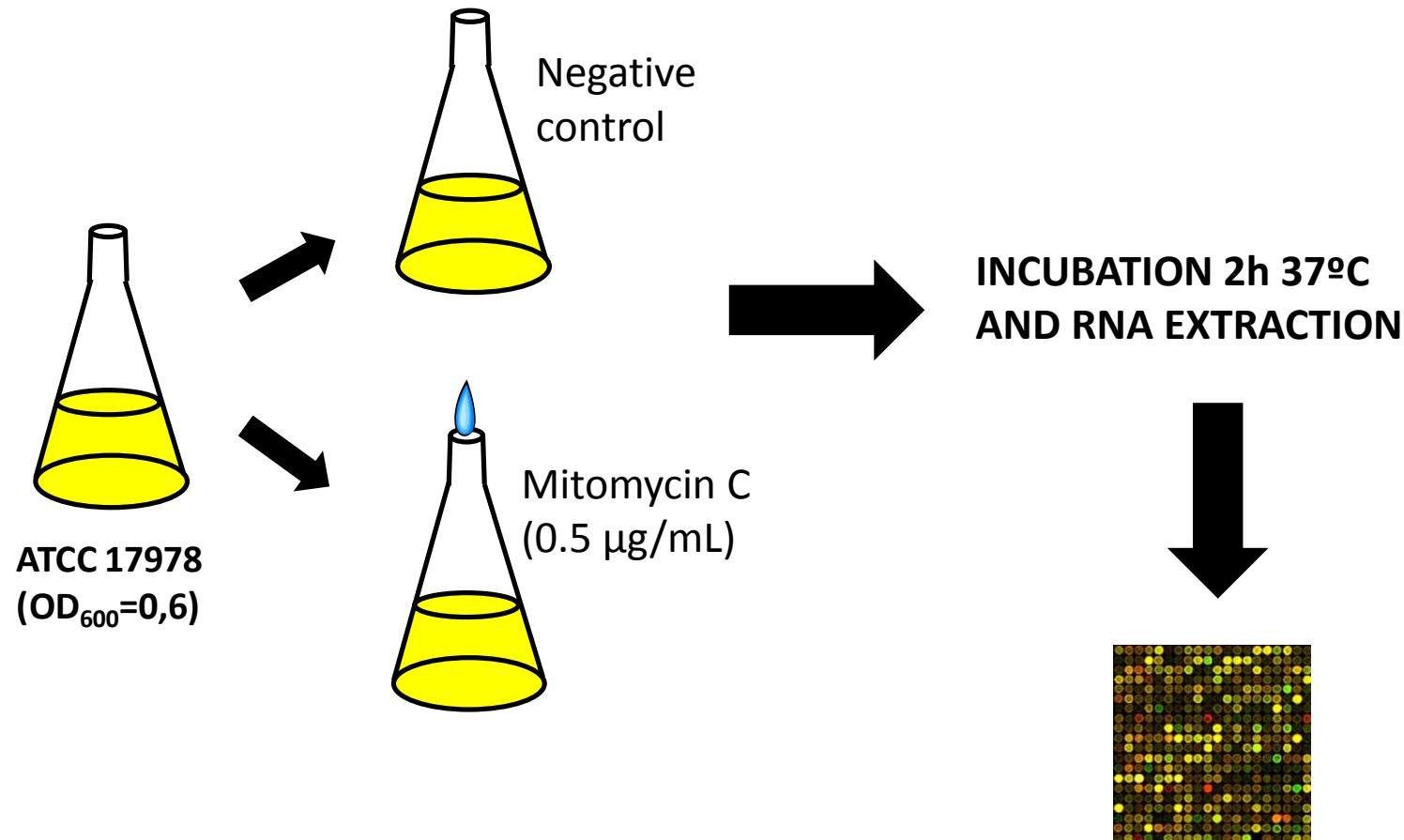
^a NA, not applicable (the pET-RA plasmid carries rifampin resistance).

^b pET-RAØ, empty pET-RA vector.

Objective II

Analysis of the transcriptional
response of *A. baumannii* to
DNA-damaging agents

Transcriptional response to MMC



Transcriptional response to MMC

Genes deregulated in MMC-treated wild-type *A. baumannii*

Predicted functional group and gene ^a	Product description ^b	Expression level (fold change) ^c	Predicted functional group and gene ^a	Product description ^b	Expression level (fold change) ^c
Prophage					
<i>AIS_1145</i>	Putative Cro protein	2.4	<i>AIS_0636</i>	DNA polymerase V component UmuD	3.7
<i>AIS_1149</i>	Putative phage-related protein	1.6	<i>AIS_1174</i>	DNA polymerase V component UmuD	2.8
<i>AIS_1155</i>	Putative phage-related protein	1.5	<i>AIS_1389</i>	DNA polymerase V component UmuDAb	2.7
<i>AIS_1156</i>	Putative phage-related protein	1.6	<i>AIS_1962</i>	Recombinase A	2.1
<i>AIS_1581</i>	Putative methyltransferase	1.9	<i>AIS_2008</i>	DNA polymerase V component UmuC	3.1
<i>AIS_1583</i>	Hypothetical protein (putative family peptidase S24)	1.5	<i>AIS_2015</i>	DNA polymerase V component	1.5
<i>AIS_1587</i>	Phage terminase EsvK2	2	<i>AIS_2035</i>	Hypothetical protein (putative endonuclease)	2
<i>AIS_1588</i>	Phage terminase-like protein large subunit	1.9	<i>AIS_2036</i>	DNA cytosine methyltransferase	1.7
<i>AIS_1590</i>	Peptidase U35 phage prohead HK97	1.9	<i>AIS_2039</i>	Hypothetical protein (DNA polymerase III subunit-like)	3.2
<i>AIS_1591</i>	Phage major capsid protein HK97	2.8	<i>AIS_2586</i>	dGTP triphosphohydrolase-like protein	2.3
<i>AIS_1592</i>	Phage head-tail adaptor	2.1	<i>AIS_3115</i>	Hypothetical protein (putative DNA metabolism protein)	2.5
<i>AIS_1593</i>	Hypothetical protein (putative phage protein HK97 gp10 family)	2.3	<i>AIS_3116</i>	Hypothetical protein (putative DNA repair SAM protein)	3.7
<i>AIS_1594</i>	Hypothetical protein (putative phage protein)	2.2	<i>AIS_3295</i>	Nucleotide excision repair component UvrA	4.3
<i>AIS_1595</i>	Hypothetical protein (phage tail protein)	1.8	Unknown		
<i>AIS_1596</i>	Hypothetical protein (phage tail protein)	1.8	<i>AIS_1143</i>	Hypothetical protein	3.4
<i>AIS_1597</i>	Lambda family phage tail tape measure protein	1.7	<i>AIS_1226</i>	Hypothetical protein	1.9
<i>AIS_1598</i>	Hypothetical protein (putative phage protein)	1.9	<i>AIS_1388</i>	Hypothetical protein DdrR	4.2
<i>AIS_1599</i>	Hypothetical protein (putative phage protein)	2	<i>AIS_2033</i>	Hypothetical protein	1.5
<i>AIS_1600</i>	Lysozyme	2.1	<i>AIS_2038</i>	Hypothetical protein (putative lipoprotein)	4.2
			<i>AIS_3385</i>	Putative membrane protein	1.9

Objective III

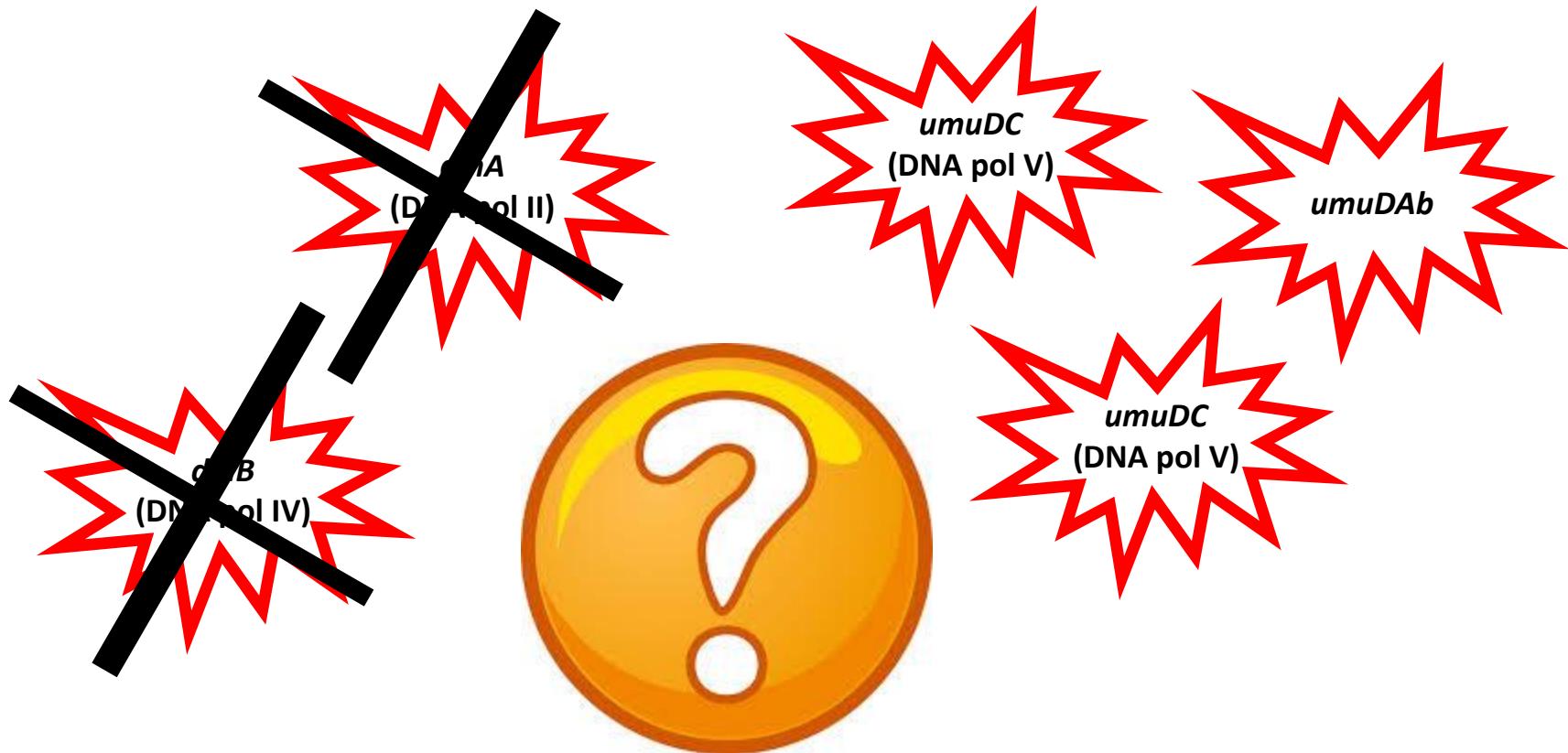
Analysis of the DNA damage-induced mutagenesis in *A. baumannii*

DNA damage-induced mutagenesis in *E. coli*

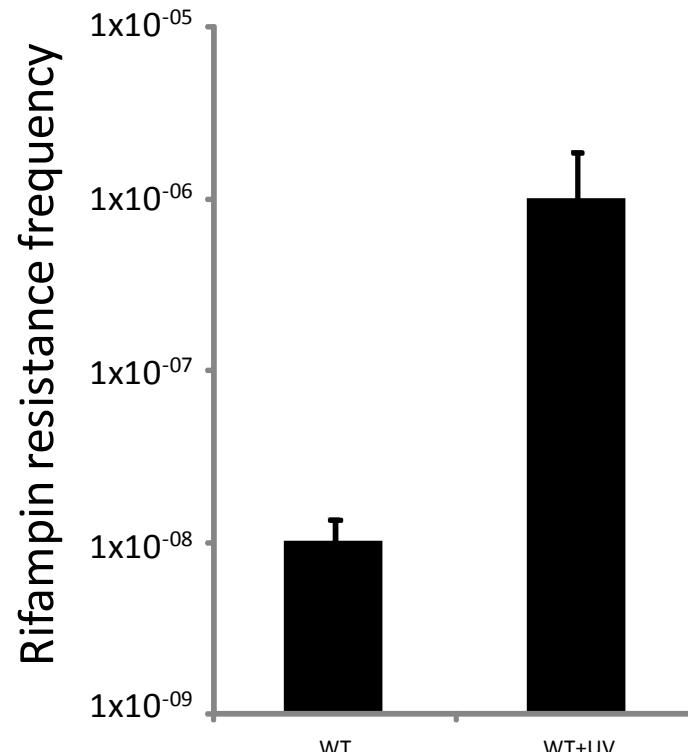


The induction of genes encoding error prone DNA polymerases causes an increase of the mutation rate

DNA damage-induced mutagenesis in *A. baumannii* ?



DNA damage-induced mutagenesis in *A. baumannii*

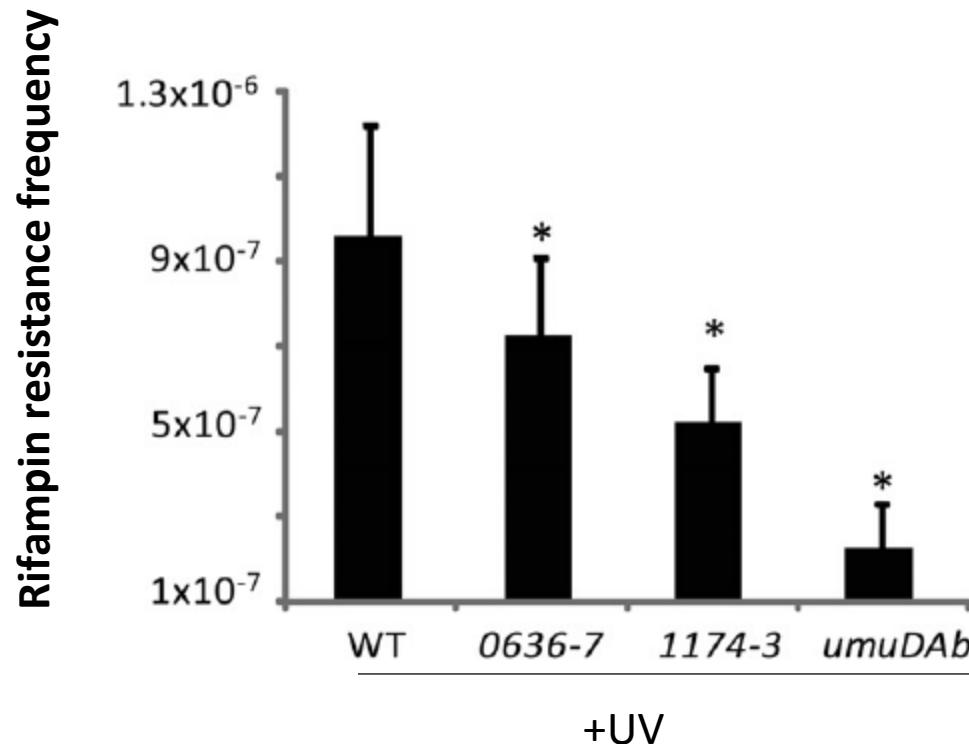


A. baumannii ATCC 17978

UV: Treatment (100 J/m²)

*P<0,01

DNA damage-induced mutagenesis in *A. baumannii*



UV: Treatment (100 J/m²)
*P<0,05

Transcriptional response to MMC

Genes deregulated in MMC-treated wild-type *A. baumannii*

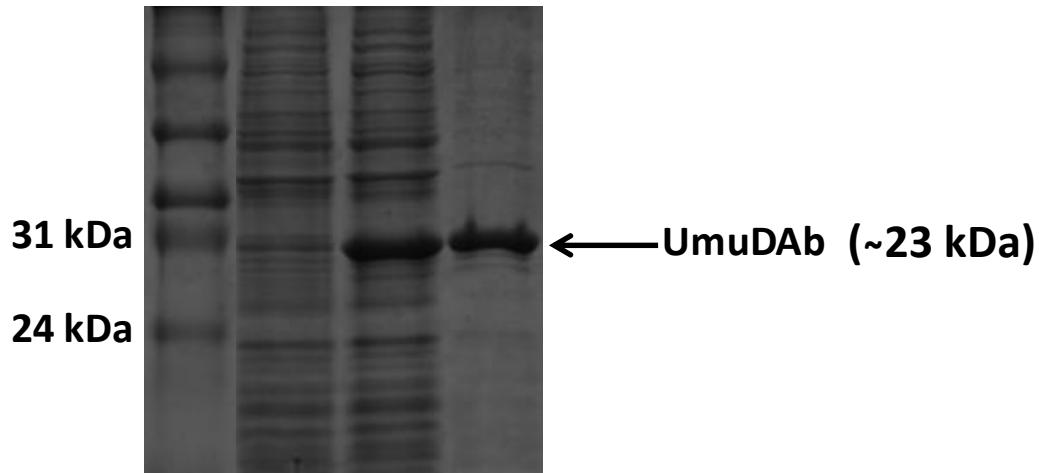
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<i>AIS_1599</i>	Hypothetical protein (putative phage protein)	2			
<i>AIS_1600</i>	Lysozyme	2.1			
DNA repair					
			<i>AIS_0636</i>	DNA polymerase V component UmuD	3.7
			<i>AIS_1174</i>	DNA polymerase V component UmuD	2.8
			<i>AIS_1389</i>	DNA polymerase V component UmuDAb	2.7
			<i>AIS_1962</i>	Recombinase A	2.1
			<i>AIS_2008</i>	DNA polymerase V component UmuC	3.1
			<i>AIS_2015</i>	DNA polymerase V component	1.5
			<i>AIS_2035</i>	Hypothetical protein (putative endonuclease)	2
			<i>AIS_2036</i>	DNA cytosine methyltransferase	1.7
			<i>AIS_2039</i>	Hypothetical protein (DNA polymerase III subunit-like)	3.2
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			<i>AIS_1226</i>	Hypothetical protein	1.9
			<i>AIS_1388</i>	Hypothetical protein DdrR	4.2
			<i>AIS_2033</i>	Hypothetical protein	1.5
			<i>AIS_2038</i>	Hypothetical protein (putative lipoprotein)	4.2
			<i>AIS_3385</i>	Putative membrane protein	1.9

UmuDAb

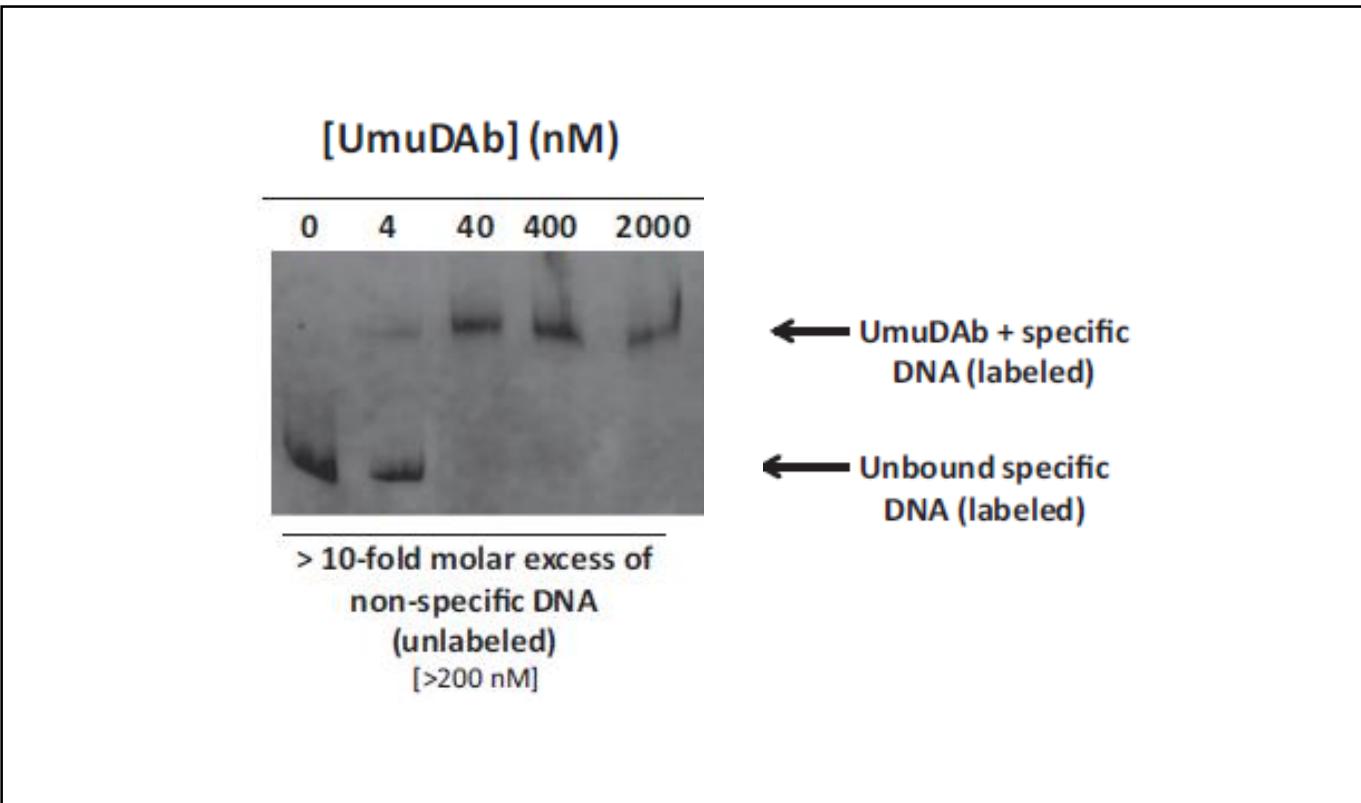
- Is present in practically all *Acinetobacter* spp.
- Encodes an extra N-terminus region

UmuDAb	MPKKKEFEHGGARENAGRKAQFNEPTKVIRVPESQVNFIKNWLLNNVKTNNQTDFTSKLK	60
UmuDE.coli	-----	MLFIKPAD 8

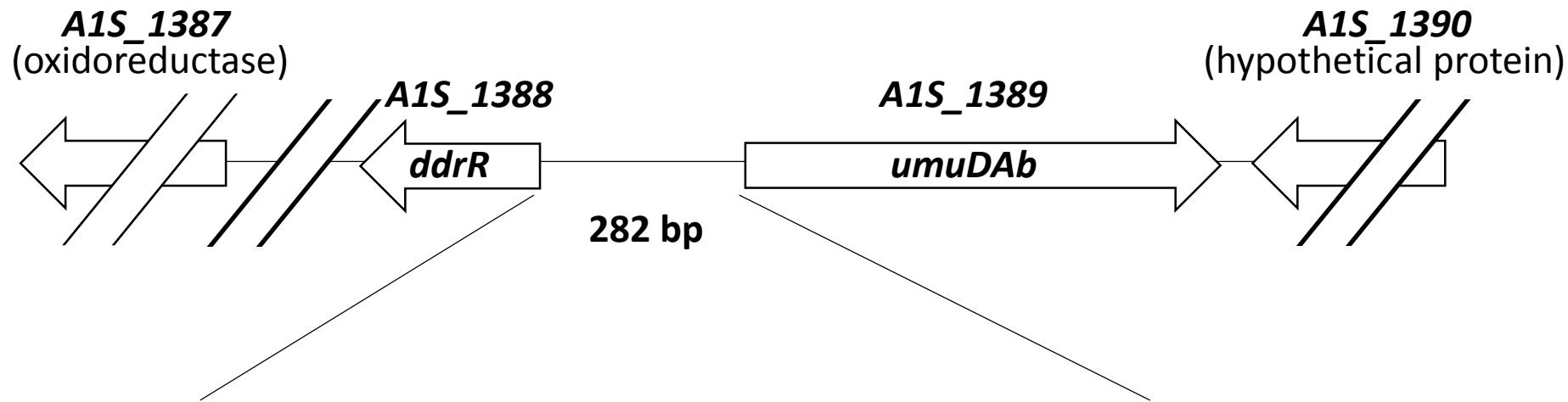
Purification of UmuDAb



UmuDAb specifically binds to its promoter region

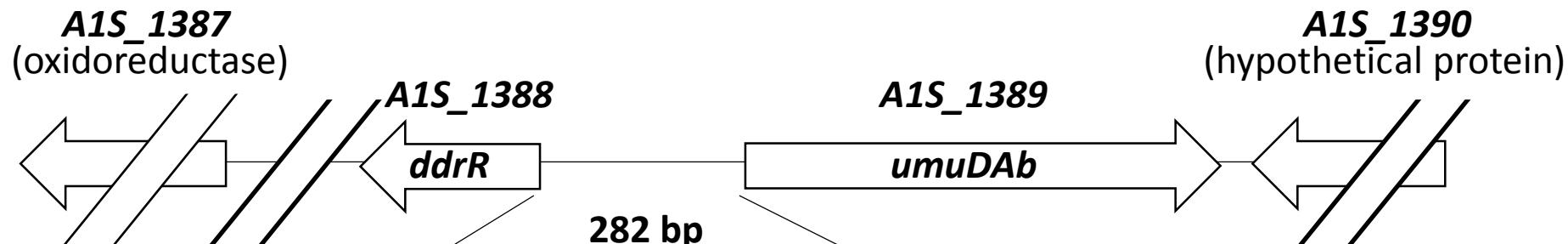


The promoter region of *umuDAb*



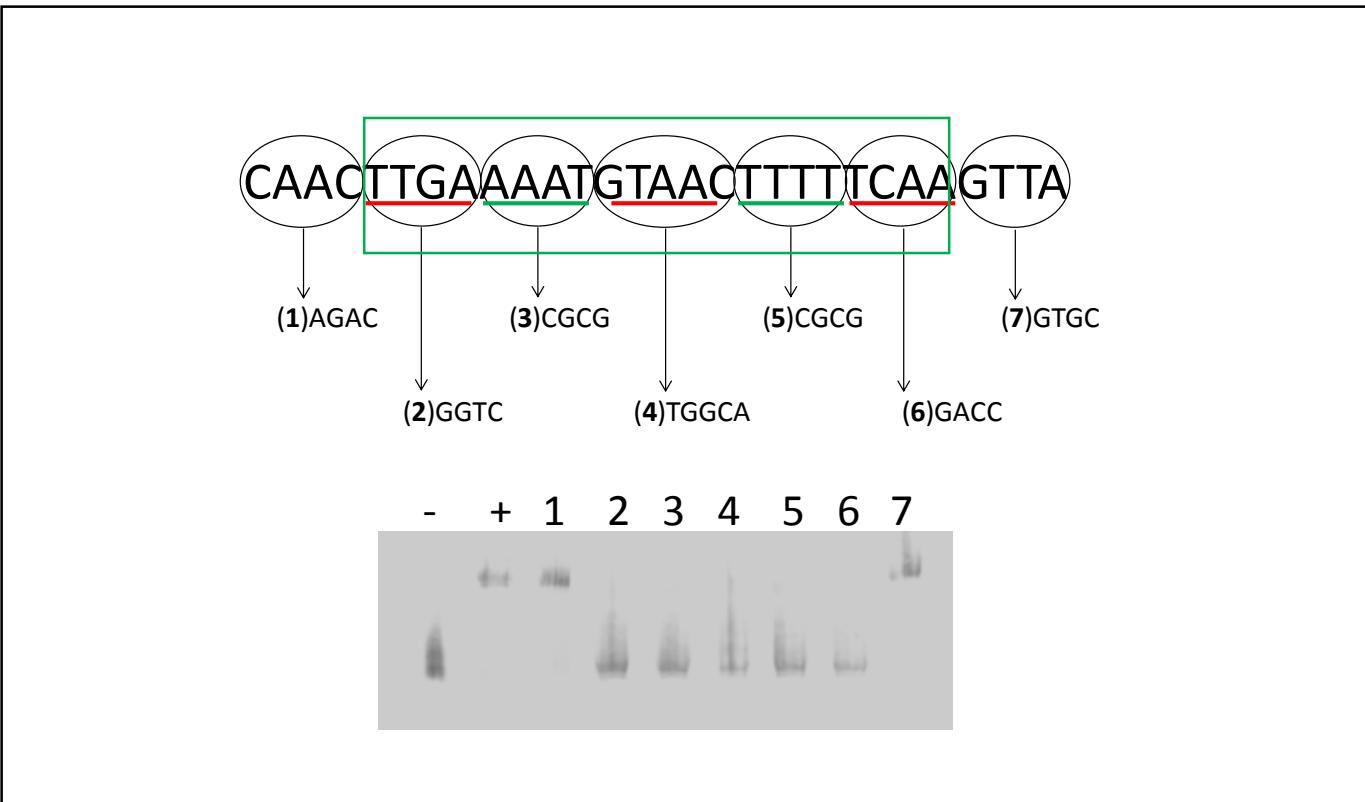
TTCGCATGTCTCCATACCAAAAAACTGAATCTAACTCATATT
AGTGTTCAGAATTTCATTGATAAAAATCAAACAGTCAGTAA
TAAGAACTATTCACAATAGGTTTGATTCTCTCAAATTACC
ATTTCCGTTAACATCCCTGTTAAAAGCAAAATGGATTAAA
ATCTGATCATGAAATTATTGTTATGCAAAAAAAATTTCGT
CAACTTGAAAATGTAACCTTTCAAGTTACATTGTTGTGA
GGATTGAAACGGTGAAATGGAGGCGAT

The promoter region of *umuDAb*



TTCGCATGTCTCCATACCAAAAAACTGAATCTAACTCATATT
AGTGTTCAGAATTTCATTGATAAAAAATCAAACAGTCAGTA
ATAAGAACTATTCACAATAGGTTTGATTCTCTTCAAATTAC
CATTCCGTTAACATCCCTGTTAAAAGCAAAATGGATTAA
AATCTGATCATGAAATTATTGTTATGCAAAAAAAATTTCG
TCAAC **TTGA** **AAAT** **GTAAC** **TTTT** **TCAA** GTTA
CATTGTTGTGTAGGATTGAACGGTGAAATGGAGGCGAT

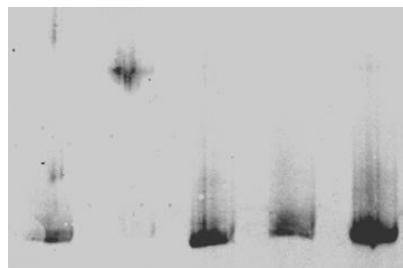
UmuDAb specifically binds to its promoter region



UmuDAb specifically binds to its promoter region

TTGA AAAT GT AAC TTTT TCAA

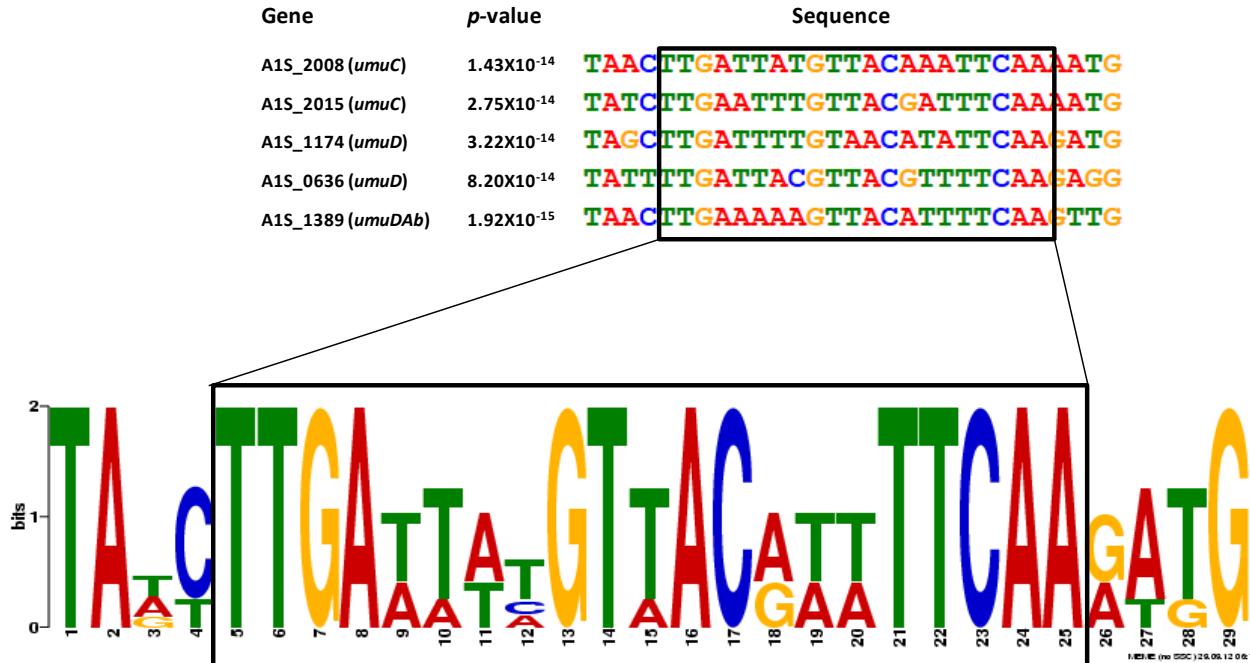
- + +C +CC +CCC



The UmuDAb regulon

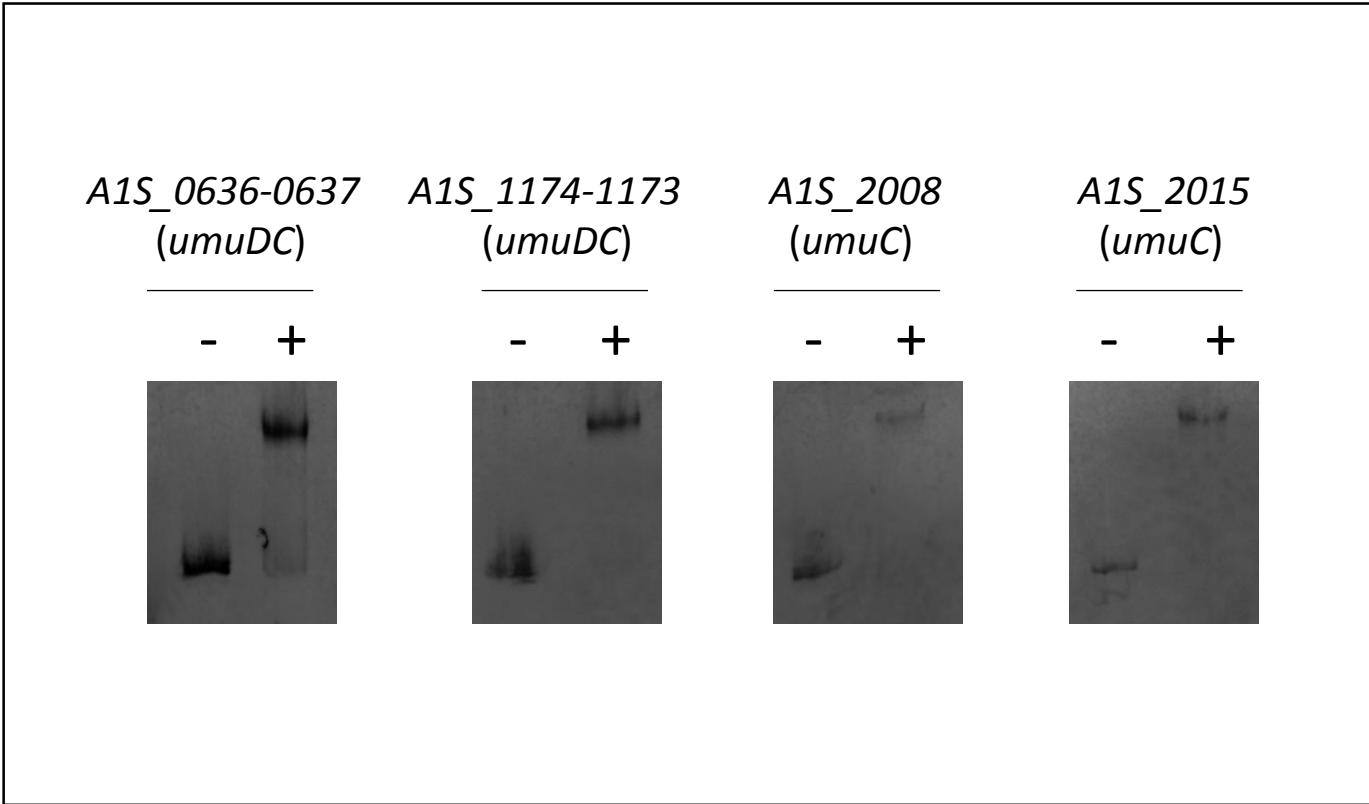
The MEME Suite

Motif-based sequence analysis tools



CONSENSUS: TTGA NNNN GTNAC NNNN TCAA

The UmuDAb regulon



The UmuDAb regulon

Inactivation of the *umuDAb* gene resulted in the deregulation of all DNA-damage-induced genes containing the described palindromic DNA motif

Gene	Designation	Product	Sequence	Position
<i>AIS_0636</i>	<i>umuD</i>	DNA polymerase V component	TTGATTACGTTACGTTTCAA	12
<i>AIS_0637</i>	<i>umuC</i>	DNA polymerase V component		
<i>AIS_1174</i>	<i>umuD</i>	DNA polymerase V component	TTGAATATGTTACAAAATCAA	15
<i>AIS_1173</i>	<i>umuC</i>	DNA polymerase V component		
<i>AIS_1389</i>	<i>umuDAb</i>	DNA polymerase V component	TTGAAAATGTAACCTTTCAA	45
<i>AIS_2008</i>	<i>umuC</i>	DNA polymerase V component	TTGATTATGTTACAAATTCAA	212
<i>AIS_2015</i>	<i>umuC</i>	DNA polymerase V component	TTGAATTGTTACGATTCAA	169

Conclusions

- I. The *A. baumannii* RecA protein is involved in repair of DNA damage, antimicrobial resistance, general stress response and virulence.
- II. *A. baumannii* lacks the LexA repressor and the error-prone DNA polymerase UmuDAb carries out its functional role.
- III. The *A. baumannii* UmuD homologs are involved in antibiotic resistance acquired through DNA damage-induced mutagenesis.